

November 2018 ISSN 1996-0778 DOI: 10.5897/AJBR www.academicjournals.org



# **ABOUT AJBR**

**The African Journal of Biochemistry Research (AJBR)** (ISSN1996-0778) is published Monthly (one volume per year) by Academic Journals.

African Journal of Biochemistry Research (AJBR) provides rapid publication (monthly) of articles in all areas of Biochemistry such as Nutritional biochemistry, Analytical biochemistry, Clinical Biochemistry, Human and Plant Genetics, Molecular and Cell Biology, Enzymology, Toxicology, Plant Biochemistry, Biochemistry Education etc. The Journal welcomes the submission of manuscripts that meet the general criteria of significance and scientific excellence. Papers will be published shortly after acceptance. All articles are peer-reviewed.

# **Contact Us**

Editorial Office:	ajbr@academicjournals.org
Help Desk:	helpdesk@academicjournals.org
Website:	http://www.academicjournals.org/journal/AJBR
Submit manuscript online	http://ms.academicjournals.me/

# Editor

## **Associate Editors**

Prof. Johnson Lin School of Biochemistry, Genetics, Microbiology and Plant Pathology University of KwaZulu-Natal (Westville) Private Bag X 54001, Durban Republic of South Africa

Gregory Lloyd Blatch Dept Biochemistry Microbilogy& Biotechnology Rhodes University Grahamstown 6140 South Africa

Dr. SerapYalin Mersin University, Faculty of Pharmacy, Department of Biochemistry, YenisehirKampusu, Mezitli 33161 Mersin/Turkey

Dr. Om Prakash Gupta Directorate of Wheat Research (ICAR) Post Box-158, A grasainMarg, Karnal-132001, Haryana, India

# **Editorial Board**

Dr. Desouky A.M. Abd-El-Haleem Biological Sciences Department, College of Arts and Sciences, Qatar University, Doha, Qatar

Dr. S.K. Trigun Biochemistry and Molecular Biology Section, Banaras Hindu University Varanasi-221005, India

Dr. ImedGallouzi McGill University, Biochemistry Department, 3655 Promenade Sir William OslerMontreal, Quebec, H3G 1Y6, Canada

Dr. Ashraf A Khalil Protein Technology Lab, Mubarak City for Science, New Borg Elarab, Alexandria, Egypt.

Dr. Stanley Mukanganyama Department of Biochemistry, University of Zimbabwe, Box MP 167, Mount Pleasant, Harare, Zimbabwe

Prof. Salah A. Sheweita Taibah University, Faculty of Medicine, Department of Biochemistry, PO Box 30001, Madinah, Saudi Arabia

Dr Oluwafemi O Oguntibeju Department of Clinical Biochemistry, School of Medicine, Spartan Health Sciences University, P.O. Box 324, Vieux Fort, St Lucia, West Indies

Dr. Robert L. Brown USDA ARS, Southern Regional Research Center 1100 Robert E. Lee Blvd., New Orleans, LA 70124 Dr. Edward Eteshola Biomedical Engineering Center Davis Heart and Lung Research Institute Ohio State University 473 W. 12th Avenue Columbus, OH 43210

G. Suresh Kumar Senor Scientist and Head Biophysical Chemistry Laboratory Indian Institute of Chemical Biology Council of Scientific and Industrial Research Jadavpur, Kolkata 700 032, India

Xu Lu Department of Biochemistry and Molecular Biology Colorado State University Fort Collins, CO 80523-1870 USA

Mohammed A.A Sarhan Dept. Biological Sciences Faculty of Science King Khalid University Saudi Arabia

MehrdadBehmanesh Department Of Genetics School Of Science P.O.Box 114-175 Tehran Iran Iran

Hans Verhagen P.o Box 1 3720 Ba Bilthoven The Netherlands Netherlands

P.K.Sumodan Post Graduate Department Of Zoology Government College Madappally India India

BalesengMoseki University Of Botswana Botswana Bhaskar C. Behera Agharkar Research Institute Plant Science Division India India

Luiz Claudio Miletti Universidade Do Estado De Santa Catarina Brasil

Oladipo Gabriel Sunday University Of Port Harcourt Port Harcourt-Nigeria Nigeria

Basiouny Ahmed El-Gamal Biochemistry Department Faculty Of Science Alexandria University Egypt

AminigoEbiokpo Rebecca University Of Port Harcourt Portharcourt-Nigeria Nigeria

JiaZeng Department Of Bioengineering Central South University Changsha Hunan 410083 P.R.China China

Adenike Kuku ObafemiAwolowo University Department Of Biochemistry Nigeria

Elsayed Hafez Genetic Engineering and Biotechnology Research Institute Egypt

Gabriella Castoria Via L. De Crecchio 7 -80138 Naples Department Of General Pathology Italy

SalwaSeddik Abdel-Latif 21 Elbatal Ahmed Abdel Aziz Elmohandesien Giza Egypt Erasto Vitus Mbugi Muhimbili University Biochemistry Department School Of Medicine India

Mohamed Rholam Université Paris7 - Denis-Diderot France

Hooi Ling Foo Universiti Putra Malaysia Malaysia

JayanthRao Biochemistry And Nutrition Cftri Mysore India

Maznah Ismail Universiti Putra Malaysia

Svetlana Lutsenko Oregon Health & Science University USA

Gabriel Ugwem Rivers State University Of Science And Technology P.M.B. 5080 Port Harcourt Nigeria

HariChhatpar Dept. Of Microbiology & Biotechnology Centre Faculty Of Science M.S.University Of Baroda Vadodara 390 002 Baroda India

MahiuddinAlamgir The University Of New South Wales Sydney Nsw-2052 Australia

Sheeja Samuel Edwin B.R Nahata College of Pharmacy & Research Centre India

William Cho Room 1305 13/F Block R Department of Clinical Oncology Queen Elizabeth Hospital 30 Gascoigne Road Kowloon Hong Kong Dr. SurainiAbd-Aziz Universiti Putra Malaysia Malaysia

Dr. Mustafa NumanBucak Lalahan Livestock Central Research Institute Lalahan Ankara Turkey

Alparslan Kadir Devrim Department Of Biochemistry Faculty of Veterinary Medicine Kafkas University 36040 Kars Turkey

Vasudev R. Thakkar Sardar Patel University Brd School of Biosciences Sardar Patel University Nagar

Prof. Emmanuel Anosike Department Of Biochemistry University Of Port Harcourt Nigeria

Dr. Usama Beshay New Bourg El-Arab City, Research Area Alexandria 21934 Egypt

Dr. Ramar Perumal Samy Department of Anatomy Yong Loo Lin School of Medicine National University of Singapore Singapore

Dr. Shin-ichi ONO Laboratory of Clinical Pharmacy College of Pharmacy, Nihon University Japan

Prof. Lawal Bilbis Biochemistry Department UsmanuDanfodiyo University Sokoto Nigeria

Dr. Adriana G. Chicco Department of Biochemistry University of Litoral, Santa Fe Argentina Prof. Zia-Ur Rahman Department Of Physiology and Pharmacology University Of Agriculture Falsalabad Pakistan

Dr. Oluwole Ariyo Allen University USA

Prof. Francisco Torrens Institut Universitari de Ciència Molecular Universitat de València Spain

Prof. Belkhodja Moulay University of Senia Oran Algeria

Dr. Hossam M Ashour Department of Microbiology and Immunology Faculty of Pharmacy, Cairo University Egypt

Dr. Fidelis Ocloo Biotechnology and Nuclear Agriculture Research Institute/GAEC Ghana

Ass. Prof. Alfonso Baldi Dept. Biochemistry, Sect. Pathology Second University of Naples, Italy

Dr. Anandh Babu Pon Velayutham Department of Human Nutrition Foods and Exercise 253 Wallace Hall Virginia Tech Blacksburg VA 24061 USA

Dr. Tapan K. Chaudhuri Department of Biochemical Engineering and Biotechnology Indian Institute of Technology Delhi, HauzKhas New Delhi-110016, India.

Dr. Rong Zhang Shenyang Pharmaceutical University China Ass. Prof. Tzong-Jih Cheng Department of Bio-Industrial Mechatronics National Taiwan University Taiwan

Dr. Zuyong Xia Department of Radiology, 1201 Welch Rd, Room P089, Stanford, CA 94301 USA

Dr. Pratap Kumar Das Indian Institute of Chemical Biology India

Dr. Vasudeo Pandharinath Zambare Advanced Enzyme Technologies Ltd India

Dr. A M Mujumdar Agharkar Research Institute India

Prof. Christine Clayton ZMBH ImNeuenheimer Feld 282 69120 Heidelberg Germany

Prof. Rekik Boul baba ESA Mateur Département des sciences et techniques de productions animales Tanzania

Dr. Farhad Mirzaei National Dairy Research Institute, NDRI Karnal India

Dr. ROUABHI Rachid Biology Department Tebessa University. Algeria

Prof. Vaclav Vetvicka University of Louisville USA Dr. Ramesh Putheti, Ph.D Research scientist Actavis Pharmaceuticals 10065 red run blvd,owings mills Blvd,Maryland.USA.21030 USA

Prof. Dr. Mustafa NAZIROGLU Head of Department of Biophysics Medical (TIP) Faculty, SuleymanDemirel University Cunur, TR-32260 Isparta TURKEY

Dr. José Luis Arias Mediano GrupolnvestigaciónFarmaciaPráctica (CTS-205) Dept. Farmacia y TecnologíaFarmacéutica Facultad de Farmacia Campus Universitario de Cartuja, s/n Universidad de Granada 18071 Granada.

Ahmed Malki, PhD Lecturer of Biochemistry and Molecular Biology Biochemistry Department Fcaulty Of Science Alexandria University Alexandria, Egypt

Dr. Alireza Seidavi (PhD) Assistant Professor of Animal and Poultry Nutrition, Department of Animal Science, College of Agriculture, Islamic Azad University, Rasht Branch, Rasht, Iran

Amani S. Awaad Professor of pharmacognosy, Chemistry Department Faculty of Sciences, King Saud University . Riyadh. KSA. P.O. Box 22452, Riyadh 11495. Saudi Arabia

Dr. Abdel-TawabMossa Environmental Toxicology Research Unit (ETRU), Pesticide Chemistry Department, National Research Centre, Dokki, Egypt Dr. Amal A. Mohamed Plant Biochemistry Department, Agriculture Division - National Research Center, 31-El-Tahrir St., Dokki, Cairo – Egypt

Dr. Anabella Gaspar Department of Biochemistry, University of Pretoria, South Africa

Dr. Anna Janecka Department of Biomolecular Chemistry, Medical University of Lodz, Mazowiecka 6/8, 92-215 Lodz, Poland

Dr. Caser Abdel Horticulture Department, Dohuk University, Iraq

Dr. David Sheehan Dept Biochemistry, University College Cork, Ireland

Dr. Dayananda Chandrappa Center for Bioenergy, Department of Life and Physical Sciences, Cooperative Research, Lincoln University, Jefferson City, USA

Dr. Elsayed Abdelaal Special Graduate Faculty, University of Guelph, Onatrio, Canada

Dr. Etienne Marbaix CELL Unit, de Duve Institute, UCL-75.41, 75 avenue Hippocrate, B-1200 Bruxelles, Belgium Dr. Gary L. Firestone Department of Molecular and Cell Biology, University of California, Berkeley, CA, 94720, USA

Dr. Henryk Zielinski Institute of Animal Reproduction and Food Research, Polish Academy of Sciences, Poland

Dr. Irshad A. Nawchoo Department of Botany, University of Kashmir, India

Dr. LuchaiButkhup Department of Biotechnology, Faculty of Technology, Mahasarakham University, Mahasarakham 44000, Thailand

Dr. LuminitaVladescu Department of Analytical Chemistry, Faculty of Chemistry, University of Bucharest, Romania

Dr. Mira Debnath School of Biochemical Engineering, Institute of Technology - Banaras Hindu University, Varanasi, India

Dr. Nilesh S. Panchal Department of Biosciences, Saurashtra University, Rajkot-360005, Gujarat. India

Dr. Rayappa A. Balikai University of Agricultural Sciences, Dharwad, Karnataka- 580 005, India Dr. SaadTayyab Institute of Biological Sciences, University of Malaya, 50603 Kuala Lumpur, Malaysia

Dr. Shijun Fu Institute of Health Sciences, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences and Shanghai Jiao Tong University School of Medicine, Shanghai, P. R. China Dr. Shiming Zhang Weis Center for Research, Geisinger Clinic, Danville, Pennsylvania, USA

Dr. Thomas Efferth Department of Pharmaceutical Biology, Institute of Pharmacy and Biochemistry, University of Mainz, Heidelberg, 55128 Mainz, Germany

# African Journal of Biochemistry Research

 Table of Contents: Volume 12 Number 10 November 2018

# ARTICLE

**Evaluation of the effect of vitamin C on caspase 9 and oxidative stress in rheumatoid arthritis patients** Amira M. Elshamy, Nagah K. Gaafar, Nadia E. ElAshwah, Ayman A. Wagih and Abeer A. Shahba



Full Length Research Paper

# Evaluation of the effect of vitamin C on caspase 9 and oxidative stress in rheumatoid arthritis patients

Amira M. Elshamy<sup>1\*</sup>, Nagah K. Gaafar<sup>1</sup>, Nadia E. ElAshwah<sup>1</sup>, Ayman A. Wagih<sup>1</sup> and Abeer A. Shahba<sup>2</sup>

<sup>1</sup>Department of Medical Biochemistry, Faculty of Medicine, Tanta University, Egypt. <sup>2</sup>Department of Internal Medicine, Faculty of Medicine, Tanta University, Egypt.

Received 8 May, 2018; Accepted 3 September, 2018

Rheumatoid arthritis (RA) is a chronic and an autoimmune disease of the joints and is widely distributed worldwide. It is characterized by alterations of the antioxidant defense system and increased free radical formation and pro-inflammatory cytokine. The aim of the present study was to evaluate the effect of vitamin C supplementation on oxidative stress biomarkers and caspase 9 level in rheumatoid arthritis patients. This study included 30 RA patients and 30 healthy subjects. Plasma levels of malondialdehyde (MDA), total antioxidant capacity (TAC), caspase 9 and 8-hydroxy-2'-deoxyguanosine (8 OHdG) were assayed as well as blood vitamin C level. These parameters were re-evaluated in RA patients after vitamin C supplementation for one month. Increased MDA and 8 OHdG levels and reduced TAC, caspase 9 and vitamin C. Levels were demonstrated in RA patients. After vitamin C supplementation, RA patients showed significant increase in TAC and vitamin C level and significant decrease in MDA and 8 OHdG levels, plasma caspase 9 level was not significantly affected after vitamin C supplementation. Increased oxidative stress and decreased apoptosis may have an important role in the pathogenesis of RA. The administration of vitamin C supplementation may help to relieve oxidative stress and enhance the antioxidant defense in these patients.

Key words: Rheumatoid arthritis, oxidative stress, apoptosis, caspase 9, vitamin C.

# INTRODUCTION

Rheumatoid arthritis (RA) is a chronic and an autoimmune disease of the joints. It affects 0.5-1% of adults worldwide. About 30% of RA patients would become disabled in the first 2-3 years without sufficient treatment. RA is characterized by alterations of the antioxidant defense system, increased free radical formation and pro-inflammatory cytokine expression at the site of inflammation (Kundu et al., 2012). Reactive oxygen species (ROS) are produced during oxidative phosphorylation. When the production of ROS exceeds the physiological level, it induces oxidative stress and damages tissue and cellular protein, lipid, and nucleic acids (Hitchon and El-Gabalawy, 2004). Large amounts of ROS have been detected in the synovial fluid and peripheral blood of RA patient .ROS production is induced by TNF- $\alpha$  stimulation. ROS can directly degrade

\*Corresponding author. E-mail: amira.elshamy@med.tanta.edu.eg.

Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u> joint cartilage, attacking its proteoglycan and inhibiting its synthesis. ROS have also been linked to mutation of p53 in RA-derived fibroblast-like synoviocytes (RA-FLS) (Sakon et al., 2003). Among the DNA constituents, guanine is highly susceptible to the oxidative damage induced by free radicals. The oxidation product, 8hydroxy-2'-deoxyguanosine (8-OHdG) is one of the most common biomarkers used to detect oxidative DNA damage. 8-OHdG is eliminated in body fluids and can be measured (Halliwell, 2000).

The damaging effect of ROS is antagonized by antioxidants. An antioxidant is any substance that is able to scavenge free radicals or inhibit the oxidation process inside the cell. Enzymatic antioxidants include superoxide dismutase (SOD), catalase (CAT), and glutathione (GSH) related enzymes (glutathione peroxidase [GPx], glutathione reductase [GR], and thioredoxin reductase). Furthermore, the non-enzymatic antioxidants includes vitamins (A, C, and E),  $\beta$ -carotene, antioxidant minerals (copper, ferritin, zinc, manganese, and selenium) and reduced glutathione (GSH) (Kalpakcioglu and Senel, 2008).

Vitamin C (ascorbate) is an essential water-soluble vitamin in humans which must be taken in diet. It acts as a cofactor for many biosynthetic enzymes required for the synthesis of amino acid-derived macromolecules, neurotransmitters and neuropeptide hormones. Also, it acts as a cofactor for many hydroxylases enzymes such as those involved in collagen biosynthesis. Moreover, vitamin C acts as an antioxidant and prevents oxidative stress-induced damage (Monfort and Wutz, 2013). RA-FLS proliferate abnormally under the stimulatory effect of many inflammatory cytokines, such as IL-1, IL-6, and TNF- α. RA-FLS are resistant to apoptosis. yielding expansion capabilities similar to tumors. This transforms FLSs from innocent mesenchymal cells to destructive aggressors. This leads to local cartilage destruction and chronic synovial inflammation. Apoptosis induction of RA-FLS is therefore suggested as a potential therapeutic approach for RA (Noss and Brenner, 2008). Apoptosis, programmed cell death, is the physiologically preferred pathway of cell death. Apoptosis is involved in a many physiological processes such as arowth. development, differentiation and immune response. Any deregulation of the apoptotic pathway may lead to the development of diseases. Apoptosis occurs through two signaling pathways: the extrinsic pathway and the intrinsic pathway. The extrinsic pathway is induced by extracellular triggers. The intrinsic pathway involves a mitochondrial pathway (Pileczki et al., 2016).

Caspases are the central executioners of apoptosis and are responsible for the morphological features of apoptosis. The extrinsic pathway is mediated by caspase-8 while the intrinsic pathway can be initiated through caspase-9. Both pathways trigger apoptosis through the cleavage of the downstream executioner proteins (caspase-3 and -7). Although caspase-9 is an intracellular protein, it was suggested that serum caspases may be noninvasive biomarkers for detection of apoptosis (Kuida, 2000).

Disease-modifying anti-rheumatic drugs (DMARDs) are mainstay of RA therapy. Methotrexate the is recommended as the first-line treatment in patients with active RA (Saag et al., 2008). However, MTX can be cytotoxic per se and further increase the degree of cell damage in RA. MTX which is used for a long time exerts cytotoxic effects on 'S phase' of the cell cycle, and inhibits cell division. MTX exerts its effects on cells by decreasing cellular antioxidant activity, and exposing cells to the unfavorable effects of ROS, eventually inducing detrimental alterations (Yulug et al., 2013). MTX markedly increases sensitivity of cells to apoptosis mediated via either death receptor or mitochondrial pathways, in part, by increasing expression of genes whose protein products play key roles in induction of apoptosis (Spurlock et al., 2011). The effects of antioxidant materials in relieving toxic effects of MTX have been intensively investigated. Vitamin C is an important agent in enhancing the antioxidant capacity of cells (Vijayprasad et al., 2014). This study therefore should be viewed as a step in assessing potential beneficial effects of the vitamin C supplementation tested on rheumatoid arthritis. We aim to use vitamin C supplementation alongside drug treatment to reduce drug dose and thus side effects of treatment.

## PATIENTS AND METHODS

## Patients

This study included 30 healthy subjects aged (25-55) years old as a control group (Group I) and 30 RA patients aged (25-55 years old) (Group IIa), these patients were given vitamin C supplementation at a dose of 500 mg twice daily for one month (vitacid 0.5 g tablets) then patients were re-evaluated again (Group IIb). RA patients were diagnosed according to the revised criteria of the American College of Rheumatology (ACR). The disease duration was 8+ 4.6 years. Patients were collected the outpatient clinics of rheumatology unit of internal medicine department, Tanta University Hospitals, Egypt. Ethical approval of this study was obtained from the Ethical Committee of Faculty of Medicine, Tanta University (approval code 30326/05/15) and written informed consents were obtained from all patients. All RA patients were under disease-modifying antirheumatic drugs (DMARDs) either as mono-therapy or in combination. The vast majority of RA patients were consuming nonsteroidal anti-inflammatory drugs (NSAIDs) on irregular basis.

#### Justification of sample size

If the sample size is too small, the study may fail to answer its research question. If the sample size is too large, the study will be more difficult and costly than necessary. So, I adjusted this sample size by the help of previous researches (the antioxidant vitamins A, C, E and selenium in the treatment of arthritis: A systematic review of randomized clinical trials) (Canter, 2007).

#### **Exclusion criteria**

Patients suffering from chronic disorders such as diabetes mellitus,

Parameter	Group I	Group II a	Group II b	F test	P value
Vitamin C mg/dl	1.65±0.23 <sup>a</sup>	1.13±0.05 <sup>b</sup>	1.58±0.07 <sup>c</sup>	117.918	< 0.001*
MDA nmol/ml	1.07±0.24 <sup>a</sup>	2.82±0.35 <sup>b</sup>	1.2±0.13 <sup>c</sup>	429.04	< 0.001*
TAC mmol/l	0.99±0.25 <sup>a</sup>	$0.86{\pm}0.04^{b}$	0.98±0.04 <sup>c</sup>	7.295	0.001*
8OHdG ng /dl	10.11±0.3 <sup>a</sup>	11.03±0.32 <sup>b</sup>	10.17±0.33 <sup>c</sup>	80.024	< 0.001*
Caspase 9 ng/ml	8.02±0.48 <sup>a</sup>	7.33±1.4 <sup>b</sup>	7.65±0.23 <sup>b</sup>	4.725	0.011

Table 1. Biochemical data of the studied groups.

Data presented as means± SD, \*P was considered significant at <0.05 .Values on the same row with different superscript differ significantly (p< 0.05) from each other. MDA: malondialdehyde. TAC: total antioxidant capacity. 8 OHdg: 8 hydroxy 2 deoxyguanosine.

thyroid dysfunction, liver or kidney diseases were excluded.

#### **Blood sampling**

Blood samples were collected from patients and controls under aseptic precautions by venipuncture; 5.0 ml in a heparinized vial for vitamin C, TAC, MDA, 8 OHdG and caspase 9 levels assessment. 1 ml of heparinized blood samples was used to measure vitamin C level (as soon as possible within 2 h), and the remaining samples were centrifuged as soon as possible at 2000 g for 10 min at 40 C. plasma samples were stored at 70°C until the time of analysis of MDA, TAC, 8 OHdG and caspase 9 levels.

#### Assay of apoptotic and DNA damage biomarkers

Plasma caspase 9 and 8 OHdG levels were measured by commercial supplied ELISA kit supplied Chongqing Biospes, Co. (Catalog No. BYEK2175, No. BYEK1218 respectively). Assay was carried out according to the manufacturers' instructions. Using the mean absorbance value for each sample, the corresponding concentration of caspase 9 and 8 OHdG were determined from the standard curve.

#### Assay of oxidative stress biomarkers

Plasma levels of both MDA and TAC were spectrophotometric assayed using commercial kit supplied by Bio-diagnostic, Egypt. The absorbance of sample and standard were measured against blank at 534 and 510 nm for MDA and TAC respectively using semiautomatic BTS-350 Biosystems spectrophotometer.

#### Assay of vitamin C level

Blood vitamin C level was assayed according to the method of Jagota and Dani using Folin-Ciocalteu reagent which reacts specifically with ascorbic acid in a broad pH range. Vitamin C standard was prepared by dissolving 10 mg of vitamin C powder in 10 ml distilled water. The sample concentrations were calculated by interplotting from the standard curve (Jagota and Dani, 1982).

### Statistics

The results for continuous variables were expressed as means± SD and one-way analysis of variance (ANOVA) were used for their analysis. The statistical significances of differences in frequencies of variants between the groups were tested using the F test. A difference was considered significant at P values less than 0.05. All statistical calculations were performed using the computer program SPSS (Statistical Package for the Social Science) version 17 for Microsoft Windows. Correlation between the variables was examined using the Pearson's correlation coefficient.

## RESULTS

Biochemical data of the studied groups are illustrated in Table 1. Plasma MDA level and 8OHdG were significantly increased in RA patients as compared to the healthy control subjects (p< 0.0001) as depicted in Figures 1 and 2 respectively .On the other hand, caspase 9 level, vitamin C level and total antioxidant capacity were significantly decreased in RA patients as compared to control subjects (p< 0.0001) as depicted in Figures 3, 4 and 5 respectively. After vitamin C supplementation, MDA level and 8OHdG level were significantly decreased as depicted in Figures 1 and 2 respectively while vitamin C level and total antioxidant capacity were significantly increased as depicted in Figures 4 and 5 respectively. On the other hand, there was no statistically significant improvement in caspase 9 level after vitamin C supplementation in RA patients (as depicted in Figure 3 and Table 2).

## DISCUSSION

Rheumatoid arthritis (RA) is an autoimmune disease characterized by cartilage and bone destruction and systemic complications. ROS have been implicated to play an important role in RA pathogenesis (Filippin 2008). The present study showed increased level of MDA in RA patients. MDA is one of the most potential biomarkers of lipid peroxidation. The rise in lipid peroxidation is due to increased ROS which tends to increase abundantly during chronic inflammation. ROS lead to cascade stimulation of the activity of mitogen activated protein kinase (MAPK) and nuclear factor-kappa B (NF-Kb) pathways and increase the inflammatory cytokines' gene expression which finally creates immune responses and causes inflammation (Srivastava and Shrivastava, 2016). In agreement with these findings, El-barbary et al. (2011); Alver et al. (2011); Datta et al. (2014); Karaman et al.

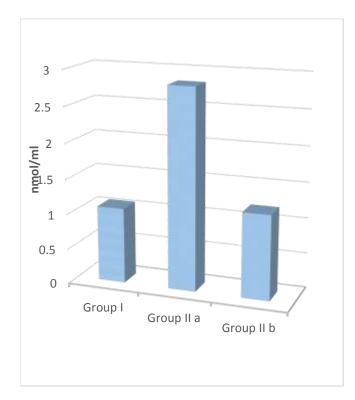


Figure 1. Statistical comparison of plasma MDA level (nmol/ml) among the studied groups using ANOVA test.

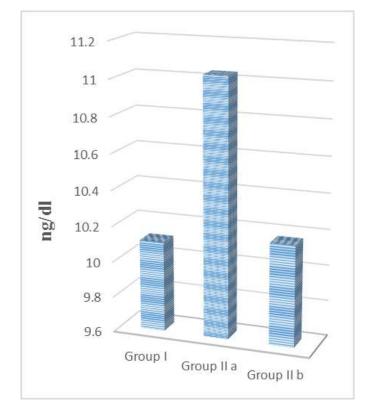
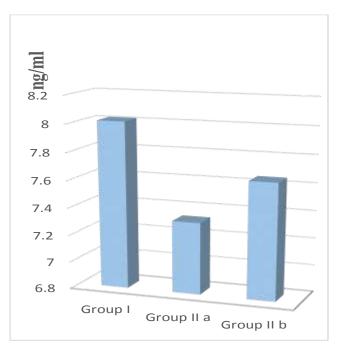
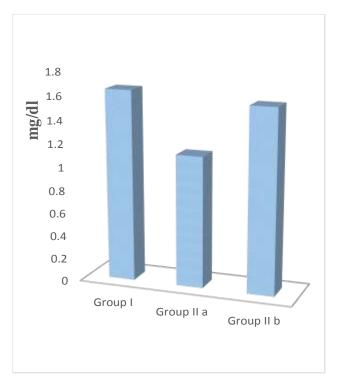


Figure 2. Statistical comparison of plasma 8 OHdG level (ng/dl) among the studied groups using ANOVA test.

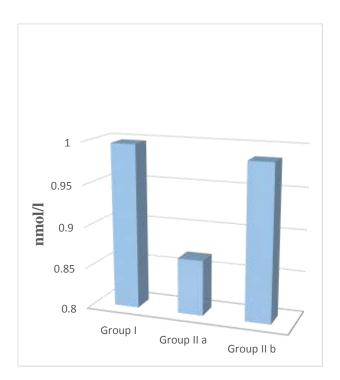


**Figure 3.** Statistical comparison of plasma caspase 9 level (ng/ml) among the studied groups using ANOVA test.



**Figure 4.** Statistical comparison of plasma vitamin C level (mg/dl) among the studied groups using ANOVA test.

(2011); Shah et al. (2011); Mishra et al. (2012) and Hassan et al. (2011) found a remarkable elevation in MDA levels in RA patients.



**Figure 5.** Statistical comparison of plasma TAC level (nmol/l) among the studied groups using ANOVA test.

		MDA	TAC	8 OHdG	Vit. C
TAC	r	-0.367			
	P-value	<0.001*			
8 OHdG	r	0.762	-0.311		
	P-value	<0.001*	0.003*		
Vit. C	r	-0.815	0.294	-0.6	
	P-value	<0.001*	0.005*	<0.001*	
Caspase 9	r	-0.253	0.178	171	0.284
	P-value	0.016	0.092	0.106	0.007*

Table 2. Spearman correlation matrix of the studied parameters.

\*P was considered significant at <0.05.

MDA: malondialdehyde. TAC: total antioxidant capacity. 8 OHdg: 8 hydroxy 2 deoxyguanosine.

On the other hand, Jacobson et al. (2012) and Kajanachumpol et al. (2000) reported no significant difference in MDA concentration between RA patients and control group. The present study showed significant elevation of plasma 8-OHdG in rheumatoid arthritis patients. 8-OHdG is the most common stable product of oxidative DNA damage. 8-OHdG is currently used as a recognized biomarkers of oxidative DNA damage mainly because of its abundance in DNA and also because of its reliable detectability (Hakem, 2008). Increased 8-OHdG level in RA patients have also been found by Kageyama et al., 2008; and Ishibashi et al., 2014; Khan et al., 2011. In contrast to the results of the present study, low level of 8-OHdG in spite of elevation of other oxidative stress biomarkers in several diseases has been reported. Sova et al. (2010) found that Serum 8-OHdG level was significantly lower in women with polycystic ovary syndrome. Inactive DNA repair enzymes may lie behind lower serum 8-OHdG levels in these patients. The main repair enzyme connected with 8-OHdG is 8-oxoguanine DNA glycosylase 1 (OGG1).With impaired OGG1 function and/or expression, cells are not able to cleave damaged guanosine from DNA, resulting in decreased 8-OHdG serum levels (Hirano, 2008).

The present study revealed significant reduction in vitamin C level and total antioxidant capacity level in RA patients. The continuous production of free radicals in inflamed joint gives rise to failure of antioxidant system and further tissue damages. Also the intake of food antioxidants is lower than the recommended amounts in RA patients (Silva et al., 2014). Reduced vitamin C level in RA patients was also reported by Jaswal et al. (2003); Mateen et al. (2016); and Vijayakumar et al. (2006). Also, Hadi et al. (2014) found significant reduction in TAC in RA. This is attributed to excess antioxidants utilization by tissues to scavenge the excessive ROS the inflamed generated at inflammatory sites. The present study found significant improvement in vitamin C level and TAC level with significant reduction of MDA and 8-OHdG levels after intake of vitamin C supplementation. Vitamin C is a chain-breaking antioxidant that halts the propagation of per-oxidative processes and reacts with membrane bound oxidized vitamin E, thus reducing it back to its native form. These findings suggest that vitamin C behaves as an ROS scavenger and may be effective in combating oxidative damage.

In agreement with these results, Meki et al. (2009) and Kuiper et al. (2011) found significant reduction in lipid peroxidation in response to vitamin C administration. Also, Al-Jassabi and Khalil (2006) found that vitamin C markedly inhibited oxidative DNA damage (as indicated by measurement of 8 OHdG). Moreover, Gęgotek et al. (2017) showed that vitamin C supplementation to fibroblasts increased GSH and SOD level and reduced MDA and total ROS levels. On the other hand, Nath et al. (2010) proved that vitamin C has no effect on 8-OHdG formation in a chemical model.

Apoptosis is a mechanism by which cells undergo programmed death. Serum markers of apoptosis may be biomarkers in patients. Moreover, noninvasive measurement of caspases in serum could be useful in monitoring different diseases (Babas et al., 2010). Owing to its crucial function of converting the death signal to the first proteolytic event and activating executioner protease, we aimed to evaluate plasma caspase 9 level in RA patients and its response to vitamin C supplementation. The present study showed significant reduction in plasma caspase 9 level in rheumatoid arthritis patients compared to control group.

Serum caspases were also evaluated in rheumatoid arthritis patients. Alzaidy et al. (2016) detected significant reduction in serum caspase3 level in rheumatoid arthritis patients. RA-FLS play an important role in both the initiation and the perpetuation of RA. RA-FLS are resistant to apoptosis. At the molecular level, synovial fibroblasts are characterized by the activation of signaling cascades that result in the inhibition of apoptosis (Khan et al., 2011). In agreement with these results, Lattuada et al. (2016) proved that the percentage of apoptotic cells (RA-FLSs) cultured in synovial fluid samples aspirated from the joints of RA was lower than in cells cultured in tissue medium alone, probably because of the presence in the SF of anti-apoptotic factors. Resistance to apoptosis in RA-FLSs depends on up regulation of IAPs which are expressed at high levels in RA-FLSs (Casnici et al., 2014). Also, Jia et al. (2015) detected increased proliferation and inhibited apoptosis in synovial cells separated from synovial tissues harvested from RA patients.

On the other hand, Yin et al. (2015) demonstrated that lipid peroxidation products induce synovial inflammations by activating NF-xB pathway and increase cleaved caspase 3 level leading to dramatic cell apoptosis. The present study revealed no significant effect of vitamin C supplementation on caspase 9 level in rheumatoid arthritis patients. This may be attributed to low dose of ascorbic acid and / short duration of treatment. Many studies proved a significant effect of vitamin C on apoptosis. Chiu et al. (2017) examined the efficacy of vitamin C to prevent monosodium iodoacetate (MIA)induced osteoarthritis in rat. In an animal model, intraarticular injection of MIA increased oxidative stress and apoptosis which resulted in cartilage degradation. All of these changes were prevented by treatment with vitamin Sato et al. (2015) demonstrated that C. Also. administration of high dose ascorbic acid reduced radiation-induced apoptosis in bone marrow cells.

On the other hand, Uetaki et al. (2015) proved that vitamin C has an apoptotic and cytotoxic effect especially on cancer cells. The administration of ascorbate (within pharmacological concentrations) is associated with the formation of  $H_2O_2$  in the extracellular fluid surrounding a tumor. In the presence of catalytic metal ions, ascorbate functions as a pro-oxidant and induces oxidative stress (Frei et al., 2008).  $H_2O_2$  can affect both extracellular and intracellular targets, as it is permeable across lipid membranes.  $H_2O_2$  may attack membrane lipids forming lipid hydro-peroxides and causing leaky membranes, intracellular oxidative stress and DNA damage leading to cell death (Du et al., 2010).

Regarding the use of intravenous vitamin C in rheumatoid arthritis, Mikirova et al. (2012) suggested that intravenous vitamin C therapy can reduce inflammation in RA patients. At high doses, ascorbate has been shown to reduce the production of pro-inflammatory cytokines and to affect the activation of NF- $\kappa$ B.

# Conclusion

Intervention with vitamin C supplementation in RA patients resulted in consistent and significant improvement of the antioxidant status and marked reduction of oxidative stress biomarkers were observed. These data are promising and indicate need for more studies to demonstrate more benefits for RA patients and to evaluate its effect on apoptosis. This study showed that vitamin C supplementation can help to relieve oxidative stress which is a serious side effect of methotrexate and cytotoxic drugs. We recommend also further studies to try to reduce the dose of these drugs with vitamin C supplementation in order to minimize side effects.

## **CONFLICT OF INTERESTS**

The authors have not declared any conflict of interests.

#### REFERENCES

- Al-Jassabi S, Khalil AM (2006). Microcystin\_induced 8\_hydroxydeoxyguanosine in DNA and its reduction by melatonin, vitamin C, and vitamin E in mice. Biochemistry (Moscow) 71(10):1115-1159
- Alver A, Şentürk A, Çakirbay H, Menteşe A, Gökmen F, Keha EE, Uçar F (2011). Carbonic anhydrase II autoantibody and oxidative stress in rheumatoid arthritis. Clinical Biochemistry 44(17-18):1385-1389.
- Alzaidy AH, Numan IT, Jassim NA (2016). Effects of adalimumab MTX combination on serum (NF-κB) and caspase 3 in Iraqi patients with rheumatoid arthritis. International Journal of Science and Research 5(7).
- Babas E, Ekonomopoulou MT, Karapidaki I, Doxakis A, Betsas G, Iakovidou-Kritsi Z (2010). Indication of participation of caspase-2 and caspase-5 in mechanisms of human cervical malignancy. IInternational Journal of Gynecological Cancer 20(8):1381-1385.
- Canter PH, Wider B, Ernst É (2007). The antioxidant vitamins A, C, E and selenium in the treatment of arthritis: a systematic review of randomized clinical trials. Rheumatology (Oxford) 46(8):1223-1233.
- Casnici C, Lattuada D, Tonna N, Crotta K, Storini C, Bianco F, Truzzi MC, Corradini C and Marelli O (2014). Optimized *in vitro* culture conditions for human rheumatoid arthritis synovial fibroblasts. Mediators of Inflammation 2014;702057.
- Chiu PR, Hu YC, Huang TC, Hsieh BS, Yeh JP, Cheng HL, Huang LW, Chang K (2017). Vitamin C protects chondrocytes against monosodium iodoacetate-Induced osteoarthritis by multiple pathways. International Journal of Molecular Sciences 18(1).
- Datta S, Kundu S, Ghosh P, De S, Ghosh A, and Chatterjee M (2014). Correlation of oxidant status with oxidative tissue damage in patients with rheumatoid arthritis. Clinical Rheumatology 33(11):1557-1564.
- Du J, Martin SM, Levine M, Wagner BA, Buettner GR, Wang SH, Taghiyev AF, Du C, Knudson CM, Cullen JJ (2010). Mechanisms of ascorbate-induced cytotoxicity in pancreatic cancer. Clinical Cancer Research 16(2):509-520.
- El-barbary AM, AbdelKhalek MA,Elsalawy AM and Hazaa SM (2011):Assessment of lipid peroxidation and antioxidant status in rheumatoid arthritis and osteoarthritis patients. The Egyptian Rheumatologist 33(4):179-185.
- Filippin LI, Vercelino R, Marroni NP, Xavier RM (2008). Redox signalling and the inflammatory response in rheumatoid arthritis. Clinical and Experimental Immunology Clinical and Experimental Immunology, 152(3):415-422.
- Frei B, Lawson S (2008). Vitamin C and cancer revisited. Proceedings of the National Academy of Sciences 105(32):11037-11038.
- Gęgotek A, Bielawska K, Biernacki M, Zaręba I, Surażyński A, Skrzydlewska E (2017). Comparison of protective effect of ascorbic acid on redox and endocannabinoid systems interactions in in vitro cultured human skin fibroblasts exposed to UV radiation and hydrogen peroxide. Archives of Dermatological Research 309(4):285-303.
- Hadi V, Kheirouri S, Alizadeh M, Khabbazi A, Hosseini H (2014). Effects of Nigella sativa oil extract on inflammatory cytokine response and

oxidative stress status in patients with rheumatoid arthritis: a randomized, double-blind, placebo-controlled clinical trial. Avicenna Journal of Phytomedicine 6(1):34-43.

- Hakem R (2008): DNA-damage repair: the good, the bad, and the ugly. EMBO Journal 27(4):589-605.
- Halliwell B (2000). Why and how should we measure oxidative DNA damage in nutritional Studies? How far have we come? American Journal of Clinical Nutrition 72:1082-108.
- Hassan SZ, Gheita TA,Kenawy SA,Fahim AT, El- Sorougy IM, Abdou MS (2011). Oxidative stress in systemic lupus erythematosus and rheumatoid arthritis patients: relationship to disease manifestations and activity. International Journal of Rheumatic Diseases 14(4):325-331.
- Hirano T (2008). Repair system of 7,8-dihydro-8-oxoguanine as a defence line against carcinogenesis. Journal of Radiation Research 49(4):329-340.
- Hitchon CA, El-Gabalawy HS (2004). Oxidation in rheumatoid arthritis. Arthritis Research and Therapy 6:265-278
- Ishibashi T, Sato B, Shibata S, Sakai T, Hara Y, Naritomi Y, Koyanagi S, Hara H, Nagao T (2014). Therapeutic efficacy of infused molecular hydrogen in saline on rheumatoid arthritis: A randomized, double-blind, placebo-controlled pilot study. International Immunopharmacology 21 (2):468-473.
- Jacobson GA, Ives SJ, Narkowicz C, Jones G (2012). Plasma glutathione peroxidase (GSH-Px) concentration is elevated in rheumatoid arthritis: a case-control study. Clinical Rheumatology 31(11):1543-1547.
- Jagota SK, Dani HM (1982). A new colorimetric technique for the estimation of vitamin C using Folin phenol reagent. Analytical Biochemistry 127(1):178-182.
- Jaswal S, Mehta HC, Sood AK, Kaur J (2003). Antioxidant status in rheumatoid arthritis and role of antioxidant therapy. Clinica Chimica Acta 338(1-2):123-129.
- Jia S, Zhang S, Yuan H, Chen N (2015). Lunasin inhibits cell proliferation via apoptosis and reduces the production of proinflammatorycytokines in cultured rheumatoid arthritis synovial fibroblasts. BioMed Research International 2015:346839.
- Kageyama Y, Takahashi M, Nagafusa T, Torikai E, Nagano A (2008). Etanercept reduces the oxidative stress marker levels in patients with rheumatoid arthritis. Rheumatology International 28(3):245-251.
- Kajanachumpol S, Vanichapuntu M, Verasertniyom O, Totemchokchyakarn K, Vatanasuk M (2000). Levels of plasma lipid peroxide products and antioxidant status in rheumatoid arthritis. Southeast Asian Journal of Tropical Medicine and Public Health 31(2):335-338.
- Kalpakcioglu B, Senel K (2008). The interrelation of glutathione reductase, catalase, glutathione peroxidase, superoxide dismutase, and glucose-6-phosphate in the pathogenesis of rheumatoid arthritis," Clinical Rheumatology 27(2):141-145.
- Karaman A, Binici DN, Meliko<sup>2</sup>glu MA (2011). Comet assay and analysis of micronucleus formation in patients with rheumatoid arthritis. Mutation Research/Genetic Toxicology and Environmental Mutagenesis 721(1):1-5.
- Khan WA, Moinuddin and Assiri AS (2011). Immunochemical studies on catechol-estrogen modified plasmid: Possible role in rheumatoid arthritis. Journal of Clinical Immunology 31(1):22-29.
- Kuida K (2000). Caspase-9. International Journal of Biochemistry and Cell Biology 32(2):121–124
- Kuiper HC, Bruno RS, Traber MG, Stevens JF (2011). Vitamin C supplementation lowers urinary levels of 4-hydroperoxy-2-nonenal metabolites in humans. Free Radical Biology and Medicine 50(7): 848-853.
- Kundu S, Ghosh P, Datta S, Ghosh A, Chattopadhyay S, Chatterjee M (2012). Oxidative stress as a potential biomarker for determining disease activity in patients with rheumatoid arthritis. Free Radical Research 46(12):1482-1489.
- Lattuada D, Gualtierotti R, Crotta K, Seneci P, Ingegnoli F, Corradini C, Viganò R, Marelli O, Casnici C (2016): Smac127 has proapoptotic and anti-inflammatory effects on rheumatoid arthritis Fibroblast-like synoviocytes. Mediators of Inflammation 2016:6905678.
- Mateen S, Moin S, Khan AQ, Zafar A, Fatima N (2016). Increased reactive oxygen species formation and oxidative stress in rheumatoid

arthritis. Plos One 11(4): e0152925.

- Meki AR, Hamed EA, Ezam KA (2009). Effect of green tea extract and vitamin c on oxidant or antioxidant Status of rheumatoid arthritis rat model. Indian Journal of Clinical Biochemistry 24(3):280-287.
- Mikirova N, Rogers AM, Casciari JJ, Taylor P (2012). Effect of high dose intravenous ascorbic acid on the level of inflammation in patients with rheumatoid arthritis. Modern Research in Inflammation 1(2):26-32.
- Mishra R, Singh A, Chandra V, Negi MP, Tripathy BC, Prakash J, Gupta V (2012). A comparative analysis of serological parameters and oxidative stress in osteoarthritis and Rheumatoid arthritis. Rheumatology International 32(8):2377-2382.
- Monfort, A, Wutz A (2013) Breathing-in epigenetic change with vitamin C. EMBO Reports 14: 337-346.
- Nath RG, Wu MY, Emami A, Chung FL (2010). Effects of epigallocatechin gallate, L-ascorbic acid, alpha-tocopherol, and dihydrolipoic acid on the formation of deoxyguanosine adducts derived from lipid peroxidation. Nutrition and Cancer 62(5):622-629.
- Noss EH, Brenner MB (2008). The role and therapeutic implications of fibroblast-like synoviocytes in inflammation and cartilage erosion in rheumatoid arthritis. Immunological Reviews 223:252-270.
- Pileczki V, Cojocneanu-Petric R, Maralani M, Neagoe IB, Sandulescu R (2016). MicroRNAs as regulators of apoptosis mechanisms in cancer. Clujul Medical 89(1):50.
- Saag KG, Teng GG, Patkar NM, Anuntiyo J, Finney C, Curtis JR, Paulus HE, Mudano A, Pisu M, Elkins-Melton M, Outman R, Allison JJ, Suarez Almazor M, Bridges SL Jr, Chatham WW, Hochberg M, MacLean C, Mikuls T, Moreland LW, O'Dell J, Turkiewicz AM, Furst DE, American College of Rheumatology (2008). American College of Rheumatology 2008 Recommendations for the use of nonbiologic and biologic disease-modifying antirheumatic drugs in rheumatoid arthritis. Arthritis and Rheumatism 59(6):762-84.
- Sakon S, Xue X, Takekawa M, Sasazuki T, Okazaki T, Kojima Y, Piao JH, Yagita H, Okumura K, Doi T, Nakano H. (2003). NF-kappaB inhibits TNF-induced accumulation of ROS that mediate prolonged MAPK activation and necrotic cell death. EMBO Journal 22:3898-3909.
- Sato T, Kinoshita M, Yamamoto T, Ito M, Nishida T, Takeuchi M4, Saitoh D, Seki S, Mukai Y (2015). Treatment of irradiated mice with high-dose ascorbic acid reduced lethality. PLoS One 10(2).

- Shah D, Wanchu A, Bhatnagar A (2011). Interaction between oxidative stress and chemokines: possible pathogenic role in systemic lupus erythematosus and rheumatoid arthritis. Immunobiology 216(9):1010-1017.
- Silva BN, Araújo ÍL, Queiroz PM, Duarte AL, Burgos MG (2014). Intake of antioxidants in patients with rheumatoid arthritis. Revista da Associação Médica Brasileira 60(6):555-559.
- Sova H, Morin-Papunen L, Puistola U, Karihtala P (2010). Distinctively low levels of serum 8-hydroxydeoxyguanosine in women with polycystic ovary syndrome. Fertility and Sterility 94(7):2670-2673.
- Spurlock CF, Aune ZT,Tossberg JT, Collins PL, Aune JP,Huston JW, Crooke PS,Olsen NJ, Aune TM (2011): Increased sensitivity to apoptosis induced by methotrexate is Mediated by Jun N-terminal kinase. Rheumatoid Arthritis 63(9):2606-2616.
- Srivastava KC, Shrivastava D (2016). Analysis of plasma lipid peroxidation and antioxidant enzymes status in patients of oral leukoplakia: A case control study. Journal of International Society of Preventive and Community Dentistry 6(3):213-218.
- Uetaki M, Tabata S, Nakasuka F, Soga T, Tomita M (2015). Metabolomic alterations in human cancer cells by vitamin C-induced oxidative stress. Scientific Reports 5:13896.
- Vijayakumar D, Suresh K, Manoharan S (2006). Lipid peroxidation and antioxidant status in blood of rheumatoid arthritis patients. Indian Journal of Clinical Biochemistry 21(1):104-108.
- Vijayprasad S, Ghongane BB, Nayak BB (2014). Effect of vitamin C on male fertility in rats subjected to forced swimming stress. Journal of Clinical and Diagnostic Research 8:5-8.
- Yin G, Wang Y, Cen XM, Yang M, Liang Y, Xie QB (2015). Lipid peroxidation-mediated inflammation promotes cell apoptosis through activation of NF- $\kappa$ B pathway in rheumatoid arthritis synovial cells. Mediators of Inflammation 2015:460310.
- Yuluğ E, Türedi S, Alver A, Türedi S, Kahraman C (2013). Effects of resveratrol on methotrexate-induced testicular damage in rats. Scientific World Journal 2013:489659.

# **Related Journals:**











www.academicjournals.org